Occurrence of antibodies to *Toxoplasma gondii* in rheas (*Rhea americana*) and ostriches (*Struthio camelus*) from farms of different Brazilian regions

Ocorrência de anticorpos *Toxoplasma gondii* em emas (*Rhea americana*) e avestruzes (*Struthio camelus*) de propriedades de diferentes regiões brasileiras

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Abstract

This study aimed to verify the occurrence of antibodies against *Toxoplasma gondii* in rheas (*Rhea americana*) and ostriches (*Struthio camelus*) commercially breeding in Brazil. Blood samples from 20 rheas and 46 ostriches (young and adults) were serologically tested using a technique known as modified agglutination test (MAT) at an initial titration of 1:16 for ostriches and 1:25 for rheas. Antibodies against *T. gondii* were found in 50% (10/20) of the rheas, with titers ranging from 1:25 to 1:6,400. The incidence of antibodies against *T. gondii* in ostriches was 17.4% (8/46) with titers ranging from 1:16 to 1:256. Birds showing titers higher than 1:200 for *T. gondii* were mainly the young ones. Therefore, rheas and ostriches may be parasitized by *T. gondii*, showing high levels of antibodies against this parasite.

Keywords: Toxoplasmosis, ratites, serology.

Resumo

Este estudo teve como objetivo verificar a presença de anticorpos contra o *Toxoplasma gondii* em emas (*Rhea americana*) e avestruzes (*Struthio camelus*) criados comercialmente no Brasil. Amostras de sangue de 20 emas e 46 avestruzes (jovens e adultos) foram testadas sorologicamente pela técnica conhecida como teste de aglutinação modificada (MAT) em titulações iniciais de 1:16 para avestruzes e 1:25 para emas. Anticorpos contra *T. gondii* foram encontrados em 50% (10/20) das emas com títulos variando de 1:25 a 1:6.400. A incidência de anticorpos contra *T. gondii* em avestruzes foi de 17,4% (8/46) com os títulos oscilando entre 1:16 e 1:256. Aves jovens foram aquelas que apresentaram títulos superiores a 1:200 para *T. gondii*. Portanto, emas e avestruzes podem ser parasitados por *T. gondii*, apresentando elevados níveis de anticorpos contra o parasita.

Palavras-chave: Toxoplasmose, ratitas, sorologia.

Toxoplasma gondii causes a parasitic disease of worldwide occurrence that has been identified in different species and is of great important as a zoonosis (MONTEIRO, 2011). Toxoplasmosis is a disease that is usually considered to be asymptomatic, because the host immune defense is able to control the infection and prevent further pathogenic damage. However, depending on the immune status (situations of poor nutrition, stress and immunosuppressive

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diseases, among others), a decline in the immunological defenses may occur and the individual might develop a clinical stage of the disease (FAGUNDES, 2009). It has been found that when humans and animals are infected by *T. gondii*, they may have fever, lymphadenopathy and anorexia (BOWMAN et al., 2010; MONTEIRO, 2011). Previous studies described occurrences of positive serology for *T. gondii* in ostriches and rheas (DUBEY et al., 2000; MAROBIN et al., 2004; CONTENTE et al., 2009; SOARES et al., 2010).

Rheas (*Rhea americana*) and ostriches (*Struthio camelus*) are ratites found in different countries, being rheas typically

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found in South America. Ostrich and rhea farming has become a growing business in Brazil, serving as an agricultural activity of national and international importance (FILHO; LUCIO, 2006). As this activity has grown, there has been a trend towards increasing health problems in these birds, as well as bird mortality and expenses with treatments (FILHO; LUCIO, 2006). These birds are believed to have a great importance in the *T. gondii* life cycle, since they might be consumed by predators such as felines (BOWMAN et al., 2010; MONTEIRO, 2011). Likewise, it is likely that ratites are involved in the epidemiology of *T. gondii* in large wild felines. Therefore, the aim of this study was to detect antibodies against *T. gondii* in *R. americana* and *S. camelus* from Brazilian commercial herds.

In this study were evaluated two rhea farms in the municipalities of Rio Rufino, state of Santa Catarina, and Santa Maria, state of Rio Grande do Sul, in southern Brazil, and three ostrich farms from the municipalities of São Paulo, state of São Paulo, São Miguel do Oeste, state of Mato Grosso and Santa Maria, state of Rio Grande do Sul. The first rhea farm had 40 birds: 21 adults aged from 3 to 7 years, and 19 rhea chicks from 4 to 6 months of age. Blood samples were collected from only 17 young and adult birds (Table 1). From the second farm, blood samples were collected from three young rheas aged one year. The ostrich blood samples were well distributed: from farm of the State University of São Paulo in the city of São Paulo, were collected from 17 adult ostriches out of a total population of 30 birds; from a commercial farm with over 5,000 birds in São Miguel do Oeste, 20 ostriches of 14 months of age; and from a farm that was just starting in the business in Santa Maria, nine samples were collected. In this farm, adult birds had been living on this farm for the last two

years (aged between 3 and 5 years), and young birds that were ten months of age at the time of sampling had been acquired at the age of three months. On all farms (rhea and ostrich), all the birds were kept on natural pasture and their diet was supplemented with commercial feed.

Blood samples (3 to 5 mL/bird) were collected using a needle (22 gauge) by means of brachial vein puncture. The samples were stored in tubes without anticoagulant, refrigerated at 10 °C, transported to the laboratory and centrifuged at 3,500 g for 10 min. Serum samples were stored at -20 °C until serological analysis for *T. gondii*.

Rhea and ostrich serum samples were assessed for antibodies against *T. gondii* by means of the modified agglutination test (MAT), accordance the methodology described by Desmonts and Remington (1980). The initial dilutions for the serum samples were 1:16 for ostriches (SOARES et al., 2010) and 1:25 for rheas (CONTENTE et al., 2009), in buffered saline solution. Thus, titers \geq 1:16 and \geq 1:25 were considered positive for ostriches and rheas, respectively. Based on this information, positive samples were further diluted in order to identify the maximum antibody titration for each bird.

Out of the 20 rhea serum samples analyzed using MAT, ten were identified as positive for *T. gondii* (50% seropositive) (Table 1). These results were higher than those found by other researchers. In a study on 74 rheas from commercial farms in the state of Rio Grande do Sul, Brazil, 8.1% were found to be positive for *T. gondii* using the hemagglutination test (MAROBIN et al., 2004). In another study on 69 rheas, 4.3% were found to showed antibodies against *T. gondii* (SOARES et al., 2010). In the present study, we found high anti-*T. gondii* titers (1:200, 1:3,200 and

Table 1. Antibodies against *Toxoplasma gondii* in rheas and ostriches on farms located in the municipalities of Rio Rufino, state of Santa Catarina; São Miguel do Oeste, state of Mato Grosso; and Santa Maria, state of Rio Grande do Sul, Brazil, according modified agglutination test (MAT).

Birds	Gender	Age	Farm location	Result/Titer	
		-	(city and state)	ity and state)	
Rhea	Female	7 years	RF/SC	Positive - 1:25	
Rhea	Female	7 years	RF/SC	Positive - 1:25	
Rhea	Female	5 years	RF/SC	Positive - 1:100	
Rhea	Female	5 years	RF/SC	Positive - 1:100	
Rhea	Female	5 years	RF/SC	Positive - 1:100	
Rhea	Male	3 years	RF/SC	Positive - 1:200	
Rhea	Female	6 months	RF/SC	Positive - 1:200	
Rhea	Male	6 months	RF/SC	Positive - 1:3,200	
Rhea	Female	6 months	RF/SC	Positive - 1:6,400	
Rhea	Male	4 months	RF/SC	Positive - 1:200	
Ostrich	Male	14 months	SMO/MT	Positive - 1:16	
Ostrich	Male	5 years	SM/RS	Positive - 1:16	
Ostrich	Female	3 years	SM/RS	Positive - 1:16	
Ostrich	Female	3 years	SM/RS	Positive - 1:16	
Ostrich	Female	3 years	SM/RS	Positive - 1:16	
Ostrich	Female	3 years	SM/RS	Positive - 1:64	
Ostrich	Female	10 months	SM/RS	Positive - 1:256	
Ostrich	Female	10 months	SM/RS	Positive - 1:256	

Rio Rufino, Santa Catarina (RF/SC); São Miguel do Oeste, Mato Grosso (SMO/MT); and Santa Maria, Rio Grande do Sul (SM/RS). Ostriches state of Sao Paulo were negative for *T. gondii*.

One of the first studies on *T. gondii* seroprevalence in which 973 ostriches were sampled, found that 2.9% were considered positive (DUBEY et al., 2000). In the current study, 17.4% of the serum samples from ostriches had differing levels of antibodies against *T. gondii*, ranging from 1:16 to 1:256. These results were similar to those reported by Contente et al. (2009) in São Paulo, Brazil, that demonstrated 14.36% of seropositive for *T. gondii*. Considering 17 samples collected in the farm of São Paulo seronegative for *T. gondii* in the current study, is demonstrated that the prevalence may depend mainly on the epidemiological situation.

The positivity for *T. gondii* in ostriches and rheas can be linked to several factors. Among these, the most important is free access by felines to environments shared by ratites like those investigated in the current study. Felines may have eliminated oocysts (the infective form) in their feces (MONTEIRO, 2011). Rheas and ostriches, along with many others (insects, worms, and small rodents) serve as intermediate hosts of *T. gondii* (RUIZ; FRENKEL, 1980). Therefore, rheas and ostriches play an important role in the epidemiology of toxoplasmosis when these birds ingest intermediate hosts infected by parasite, since they can be latter consumed by wild felines, infecting this predator (DUBEY; BEATTIE, 1988; MONTEIRO, 2011).

As previously mentioned, ostriches and rheas apparently showed no clinical changes, although young birds showed higher levels of circulating antibodies. Similar to ratites, toxoplasmosis in the intermediate host of backyard chickens (*Gallus gallus domesticus*) is usually asymptomatic (GARCIA et al., 2000). However, clinical signs have been recorded in some birds, such that eye and brain injuries have been seen to affect turkeys and canaries (QUIST et al., 1995; WILLIAMS et al., 2001). It has been found that implementing good sanitary management for these birds is important in order to prevent infection and environmental contamination. Thus, feed and water that are free from *T. gondii* should be provided, so as to decrease the prevalence of birds infected by the parasite, consequently reducing the risk of infection among humans (CONTENTE et al., 2009).

Based on these results, we conclude that the occurrence of *T. gondii* infection in *R. americana* and *S. camelus* varied among the farms investigated in Brazil. This study suggests that the young birds tested probably had been affected by recent infection, given that they presented higher levels of antibodies against the parasite.

Ethical approval

The procedure used was approved by the Animal Welfare Committee of Universidade do Estado de Santa Catarina (UDESC), under number 1.50.12.

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