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Cryptosporidium occurrence in ruminants from the North Pioneer mesoregion of Paraná, Brazil

Ocorrência de Cryptosporidium em ruminantes da mesorregião norte pioneiro do Estado do Paraná, Brasil

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Abstract

The aim of this study was to investigate the occurrence of *Cryptosporidium* in cattle and sheep from the North Pioneer mesoregion of the state of Paraná. For this, 317 stool samples were collected from cattle and sheep on 16 properties in six municipalities in the North Pioneer mesoregion of Paraná. For detection of *Cryptosporidium* species, molecular analysis was performed using nested-PCR techniques targeting the 18S rRNA gene. Of the 37 beef cows and 115 calves analyzed, four (10.8%) and 14 (12.2%), respectively, were positive for *Cryptosporidium*. Of the 12 cows and 52 calves, one (8.3%) and 14 (26.9%), respectively, were positive for *Cryptosporidium*; and of the 42 ewes and 59 lambs, six (14.3%) and 12 (20.3%), respectively were positive for *Cryptosporidium*. Cattle (15.3%) and sheep (17.8%) were both susceptible to infection. All the properties of the municipalities of Assaí, Ibaiti and, Leópolis presented infected animals. The study showed that *Cryptosporidium* occurs in most municipalities assessed, that dairy calves had a higher risk (Odds Ratio=2,66, *p-value*=0,018) for infection than beef calves, and that sheep are just as susceptible to infection as are cattle, and that further *Cryptosporidium* studies are developed.

Keywords: Cryptosporidiosis, cattle, sheep, occurrence, Southern Brazil.

Resumo

O objetivo deste estudo foi investigar a ocorrência de *Cryptosporidium* em bovinos e ovinos da mesorregião norte pioneiro do Estado do Paraná. Para tanto, 317 amostras de fezes destes ruminantes foram colhidas de 16 propriedades de seis municípios do Norte Pioneiro do Paraná. Para detecção de *Cryptosporidium* spp foi realizada análise molecular pela Técnica de nested-PCR direcionada ao gene 18S rRNA. Das 37 vacas de corte e 115 bezerros de corte analisados, quatro (10,8%) e 14 (12,2%) foram respectivamente positivos para *Cryptosporidium*. Das 12 vacas e 52 bezerros de leite, um (8,3%) e 14 (26,9%) foram positivos para *Cryptosporidium*, respectivamente. Bovinos (15,3%) e ovinos (17,8%) foram igualmente suscetíveis à infecção. Todas as propriedades dos municípios de Assaí, Ibaiti e Leópolis apresentaram animais infectados. Este estudo demonstrou que *Cryptosporidium* ocorre na maioria dos municípios avaliados, sendo que os bezerros de leite apresentam maior risco (Razão de chances=2,66, *p-value*=0,018) à infecção que os bezerros de corte e que os ovinos são tão suscetíveis à infecção quanto os bovinos e por isso, estudos nesta espécie animal devem ser mais desenvolvidos.

Palavras-chave: Cryptosporidiose, bovinos, ovinos, ocorrência, Sul do Brasil.

Cryptosporidium is a genus of apicomplexan protozoan from the class Conoidasida, order Eucoccidiorida, and family Cryptosporidiidae; *Cryptosporidium* species have a life cycle that

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In ruminants, the occurrence of *Cryptosporidium* has been linked to gastroenteritis in calves between 0 and 2 months of age and a high morbidity rate in dairy cattle (COKLIN et al., 2007; SANTÍN et al., 2008), calves prior to weaning and in newly weaned



calves (AYINMODE et al., 2010; FAYER et al., 2008), and in older cattle (KVÁC & VITOVEC, 2003; ROBINSON et al., 2006). In sheep, the highest infection rates are observed in the first few weeks of life (MARTINEZ & BELDA, 2001), with increased infection rates up to 21 days (SANTÍN et al., 2007). Older lambs demonstrate delayed growth, and adults are generally refractory to infection and illness (XIAO et al., 1993), but can eliminate oocysts in the feces, thereby contaminating the environment (MAJEWSKA et al., 2000; ZUCATTO et al., 2015).

In Brazil, studies on cryptosporidiosis in ruminants have primarily been performed in the Southeast, mainly in the states of Rio de Janeiro (COSENDEY et al., 2008; FIUZA et al., 2011), Minas Gerais (GARCIA & LIMA, 1993; GARCIA & LIMA, 1994; LIMA et al., 2013), and São Paulo (AQUINO et al., 2015; COELHO et al., 2016; FEITOSA et al., 2004; FÉRES et al., 2009; PAZ E SILVA et al., 2013, 2014; SEVÁ et al., 2010; SILVA-JUNIOR et al., 2011). In the state of Paraná, infection with Cryptosporidium was evaluated in captive birds (NAKAMURA et al., 2009), dogs with diarrhea (NAVARRO et al., 1997), horses (FUJII et al., 2014), wild birds and mammals (SNAK et al., 2015), cattle (TOLEDO et al., 2017) and sheep (SNAK et al., 2017). However, epidemiological studies have not been conducted on cryptosporidiosis in ruminants of the north pioneer mesoregion of Paraná. Knowledge about cryptosporidiosis among ruminants in this region of Paraná may have important implications in the field of veterinary public health, since this disease is a zoonosis. When results of this type of study are published, health surveillance agencies and other health professionals can alert the local population about hygiene measures necessary for the prophylaxis of the disease in animals and people living in the area. Thus, the present study aimed to evaluate Cryptosporidium infection and to determine its frequency of occurrence among adult and young cattle, dairy cows and adult and young sheep of the region.

Fecal samples of 317 ruminants who were apparently healthy, without symptoms of diarrhea, from 16 farms were chosen at random from six municipalities of North Pioneer, as follows: 75 samples from two farms in the municipality of Assaí, 63 samples from six farms in the municipality of Santo Antonio da Platina, 44 samples from two farms in the municipality of Ibaiti, 48 samples from three farms in the municipality of Ribeirão do Pinhal, 20 samples from one farm in the municipality of Ribeirão Claro, and 67 samples from two farms in the municipality of Leópolis. The period of sample collection was from August 2013 to May 2014. Of the 317 samples collected, 115 were from beef calves (aged \leq 7 months), 52 from dairy calves (aged \leq 6 months), 37 beef cows (aged > 3 years), 12 dairy cows (aged > 3 years), 59 from lambs (aged \leq 4 months), and 42 from ewes (aged > 1.5 years). The collected samples were transported in a refrigerated condition to the Laboratory of Parasitology and Parasitic Diseases of the State University of North of Paraná; and an aliquot of each sample was separated and frozen until DNA extraction.

DNA extraction was performed using a commercial kit— NucleoSpin[®] Tissue (Macherey-Nagel GmbH & Co.)—and following the manufacturer's protocol, with three incremental freeze and thaw cycles before the lysis step in order to improve the rupture of oocysts (WELLS et al., 2015).

For the detection of *Cryptosporidium* species, a 826-840 bp fragment of the 18S ribosomal RNA (rRNA) gene was amplified

by nested polymerase chain reaction (PCR) with primers (XIAO et al., 1999).

Negative controls consisting of ultrapure water and positive controls consisting of *C. parvum* DNA were used in all batches. The products were subjected to 1.5% agarose gel electrophoresis, stained with SYBR Safe[®] (Invitrogen Co.), and photographically documented.

A descriptive variable analysis (positive/negative) was performed for *Cryptosporidium* infection in the populations of cattle and sheep, as well as for determining the odds ratio and their respective confidence intervals for the exposure variable (age, type, and category) and outcome (infection). The association between risks was assessed using Fisher's exact test, with significance set at *p-value*< 0.05. Statistical analyses were performed using GraphPad Prism 6.01 (GraphPad Software, Inc.).

This project was approved (CEUA 3164-48) by the Comitê de Ética do Uso de Animais (CEUA), Universidade Estadual do Norte do Paraná.

Of the total 317 samples tested, 51 (16.1%) were positive for *Cryptosporidium*. Of the 37 beef cows and 115 calves analyzed, four (10.8%) and 14 (12.2%), respectively, were positive for *Cryptosporidium*. Of the 12 cows and 52 calves analyzed, one (8.3%) and 14 (26.9%), respectively, were positive for *Cryptosporidium*; and of the 42 ewes and 59 lambs, six (14.3%) and 12 (20.3%), respectively, were positive for *Cryptosporidium*. However, statistically significant differences were found only between beef cattle and dairy cattle. These differences were related to higher infection rates among dairy calves (26.9%) than among beef calves (12.2%), *p-value*=0.018. Dairy calves were 2.7 times more likely to be infected with *Cryptosporidium* that were beef calves (OR = 2.66; CI 1.16-6.10%)

A similar infection rate (28.1%) was reported in dairy calves in Canada by using the Sheather's flotation method (COKLIN et al., 2007), but other authors have found higher occurrence rates of *Cryptosporidium* (43.6–82.1%) using the modified Ziehl-Neelsen method (EDERLI et al., 2004), and Sheather's or Ziehl-Neelsen methods followed by PCR techniques (COUTO et al., 2014; TOLEDO et al., 2017). Of the 216 cattle evaluated, 33 (15.3%) were positive for *Cryptosporidium*, as were 18 (17.8%) of the 101 sheep. Although the rate of infection was higher in sheep, this difference was not significant (*p-value* = 0.57) (Table 1). Contrary to the results of this study, Villacorta et al. (1991) analyzed 141 cattle and 69 sheep and found infection rates of 6.38% and 1.45% respectively, using Ritchie's Technique modified by Allen & Ridley (1970).

Although not significant (*p-value* = 1.15) there were more positive lambs (20.3%) than beef calves (12.2%). While comparative studies are scarce, an earlier study of 460 sheep in Araçatuba, São Paulo showed a 6.7% positive rate for oocysts using Sheather's flotation method, which is in agreement with our findings (FÉRES et al., 2009). In the same city, 12.4% of the calves were found to be positive using this same technique in a similar number of samples (FEITOSA et al., 2004). In the State of Paraná, recent research in the Western region showed high positivity in adult sheep (59%), however the authors evaluated few lambs (7/144) (SNAK et al., 2017)

The infection rate among lambs was 43% higher than that seen in sheep. Although this difference was not significant, these

Table 1. Distribution of positive and negative animals, relative frequency (%), and statistical analyzes (OR - Odds Ratio, CI - Confidence Interval, p-value and χ^2 -Chi-square) of results from fecal tests for *Cryptosporidium* infection in adult and young cattle and sheep in the North Pioneer mesoregion of Paraná, Brazil.

Variable Animal category	Infection results					2
	Positive	Negative	- % Relative	OR (CI)	p-value	χ^2
Beef cow	4	33	10.8	1.33 (0.13-13.2)	0.805	0.06
Dairy cow	1	11	8.3			
Species						
Cattle	33	183	15.3	1.20 (0.44-1.56)	0.57	0.33
Sheep	18	83	17.8			
Age						
Young	40	186	17.7	1.56 (0.76-3.2)	0.22	1.51
Adult	11	80	12.1			
Category/Specie/Age						
Lamb	12	47	20.3	1.53 (0.52-4.47)	0.43	0.61
Ewe	6	36	14.3			
Calf	28	139	16.8	1.77 (0.65-4.87)	0.26	1.26
Cow	5	44	10.2			
Beef calf	14	101	12.2	2.66* (1.16-6.10)	0.018	5.58
Dairy calf	14	38	26.9			
Beef cow	4	33	10.8	1.33 (0.13-13.24)	0.8	0.06
Dairy cow	1	11	8.3			
Beef calf	14	101	12.2	1.84 (0.79-4.29)	1.15	2.05
Lamb	12	47	20.3			
Dairy calf	14	38	26.9	1.44 (0.59-3.48)	0.65	0.21
Lamb	12	47	20.3	. ,		
Beef cow	4	33	10.8	1.38 (0.35-5.31)	0.64	0.22
Ewe	6	36	14.3			
Dairy cow	1	11	8.3	1.83 (0.19-16.92)	0.59	0.29
Ewe	6	36	14.3	()		

*Significate difference (p < 0.05)

results were similar to those published by Causapé et al. (2002) and Paz & Silva et al. (2014).

The variation in the infection rate, as determined by the presence of oocysts in the feces of animals infected with *Cryptosporidium*, is probably because of the form of laboratory diagnosis (for direct or molecular parasitological techniques) and also the clinical condition of the animals.

Techniques such as the modified Ziehl-Neelsen method, direct fluorescent antibody test, negative staining method or ELISA, with or without subsequent PCR, are widely used in the study of cryptosporidosis in ruminants. Among these techniques, PCR has the highest sensitivity (MORGAN et al., 1998; REKHA et al., 2016; TAHVILDAR-BIDEROUNI & SALEHI, 2014); moreover, the reported rates of occurrence of *Cryptosporidium* vary between studies (CAUSAPÉ et al., 2002; EDERLI, et al., 2004; FÉRES et al., 2009; PAZ & SILVA et al., 2014; SNAK et al., 2017; TOLEDO et al., 2017). Higher infection rates are reported with the use of a stool oocyst-concentration technique, such as Sheather's flotation method (COKLIN et al., 2007; COUTO et al., 2014), probably due to its high sensitivity (REKHA et al., 2016). The discrepancy of sensitivity and specificity between the diagnostic techniques for *Cryptosporidium* occurrence indicates the need for multiple laboratory techniques for better reliability of the results.

Table 2. Distribution of the total Cryptosporidium infection rates of beef calves, dairy calves, beef cows, dairy cows, lambs, and ewes for the
municipalities of Assaí, Santo Antônio da Platina (Sto A Platina), Ibaiti, Ribeirão do Pinhal (Rib Pinhal), Ribeirão Claro (Rib Claro), and
Leópolis in the state of Paraná, Brazil.
Dairy calves provide a construction of the con

Municipalities	Total (%)	Beef calves (%)	Dairy calves (%)	Beef cows (%)	Dairy cows (%)	Lambs (%)	Ewes (%)
Assai	18.7	16.0	21.4	14.3	0	33.3	—
Sto A Platina	11.1	18.8	11.1	0	—	11.1	9.1
Ibaiti	36.4	33.3	46.7	33.3	16.7	—	37.5
Rib Pinhal	10.4	8.7		14.3	_	0	18.2
Rib Claro	0	0		0	—	0	—
Leópolis	13.4	8.0		0		29.2	0

The present study did not used diarrheic samples. This may be one reason the infection rate found in this study was lower than that reported by several other authors, including Causapé et al. (2002), that analyzed 1-week-old lambs with diarrhea and found that 93.3% of the animals were positive for Cryptosporidium infection, as determined by the Ziehl-Neelsen technique. Ayinmode et al. (2010) reported that 52.3% of PCR samples from calves with diarrhea in Nigeria were positive for Cryptosporidium infection and Garcia & Lima (1994) found almost 60% of the calves positive for Cryptosporidium infection as per the Ziehl-Neelsen technique had diarrhea. Other studies in ruminants-regardless of the presence of diarrhea—have shown infection rates lower (0-12.4%) than those reported in the present study (COKLIN et al., 2007; FEITOSA et al., 2004; FÉRES et al., 2009; CARDOSO et al., 2008).

Young animals are expected to have a higher risk for infection, as compared to the adults; however, our results indicated no difference in the occurrence of Cryptosporidium between calves (16.8%) and cows (10.2%), possibly due to the similar infection rates observed in calves and cows (12.2 and 10.8%, respectively) or the low number of cows analyzed (n = 12). Beef cows and calves live in the same environment, which possibly accounts for the similar infection rates between these sub-groups. Genotypic analyses would be necessary to confirm this hypothesis, since distinct species of Cryptosporidium infect animals of different ages. Dairy calves are separated from their mothers, and usually maintained under unhygienic conditions, which could explain the trend toward higher occurrence rates in calves than in milk cows. In previous comparative studies, Coklin et al. (2007) evaluated 143 dairy cows and found an infection rate of zero (0%) in cows and 39.6% in calves. Cardoso et al. (2008) reported Cryptosporidium infection rates of 1.8% in calves (aged 0-6 months) vs. 0.35% in cows and heifers; and Gow & Waldner (2006) reported 1.1% (5/560) in beef cows and 3.1% (19/605) in beef calves.

The similar rate of occurrence of Cryptosporidium in young and adult animals found in this study could also be explained by a high infection rate in adult cattle by C. andersoni. This species is found in animals from 1 year of age to older animals (up to 2 years old), often asymptomatic bearers of environmental contamination (ROBINSON et al., 2006)

Cryptosporidium infections in beef cattle are not frequent (HECKLER et al., 2015). In Brazil, Cardoso et al. (2008) found only 0.4% of cows to be positive for Cryptosporidium in the municipality of Caçapava, São Paulo. Ralston et al. (2003), and Gow & Waldner (2006) also identified a low prevalence of Cryptosporidium in calves and were among the few researchers to evaluate beef cattle.

Of the 16 properties studied, 11 (68.7%) had at least one animal that was positive for *Cryptosporidium*. All properties in the municipalities of Assaí (2/2), Ibaiti (2/2), and Leópolis (2/2) had infected animals. Half of the Santo Antônio da Platina properties (50%) and 66.7% of the Ribeirão do Pinhal properties had positive animals (Table 2). Epidemiological studies of parasitic diseases in the North Pioneer mesoregion are scarce. However, the few studies on ruminants in this region have shown high parasite loads, which may reflect poor sanitary conditions and inadequate handling of these animals (HOLSBACK et al., 2013, 2015, 2016).

It was concluded that Cryptosporidium occurs in most municipalities assessed, that dairy calves have a higher risk for infection than beef calves, and that sheep are just as susceptible to infection as cattle. This was the first epidemiological study of cryptosporidiosis in ruminants in the North Pioneer mesoregion of Paraná.

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References

Allen AVH, Ridley DS. Further observations on the formol-ether concentration technique for faecal parasites. J Clin Pathol 1970; 23(6): 545-546. http://dx.doi.org/10.1136/jcp.23.6.545. PMid:5529256.

Aquino MCC, Widmer G, Zucatto AS, Viol MA, Inácio SV, Nakamura AA, et al. First molecular characterization of Cryptosporidium spp. infecting buffalo calves in Brazil. J Eukaryot Microbiol 2015; 62(5): 657-661. http:// dx.doi.org/10.1111/jeu.12223. PMid:25941018.

Ayinmode AB, Olakunle FB, Xiao L. Molecular characterization of Cryptosporidium spp. in native calves in Nigeria. Parasitol Res 2010; 107(4): 1019-1021. http://dx.doi.org/10.1007/s00436-010-1972-1. PMid:20644959.

Cardoso JMS, Silveira FL, Araújo AJUS, Carvalho JCC, Kanamura HY. Ocorrência de Cryptosporidium spp. em um rebanho bovino leiteiro no município de Caçapava, estado de São Paulo, Brasil. Rev Bras Parasitol Vet 2008; 17(Supl S1): 239-242.

Causapé AC, Quílez J, Sánchez-Acedo C, del Cacho E, López-Bernad F. Prevalence and analysis of potential risk factors for *Cryptosporidium parvum* infection in lambs in Zaragoza (northeastern Spain). *Vet Parasitol* 2002; 104(4): 287-298. http://dx.doi.org/10.1016/S0304-4017(01)00639-2. PMid:11836029.

Clode PL, Koh WH, Thompson RCA. Life without a host cell: what is *Cryptosporidium? Trends Parasitol* 2015; 31(12): 614-624. http://dx.doi. org/10.1016/j.pt.2015.08.005. PMid:26440789.

Coelho WMD, Widmer G, Oliveira FP, Amarante AFT, Bresciani KDS. Natural infection by *Cryptosporidium* spp. in production animals: first description of subtype IIaA15G2R1 in goat kids and piglets in Brazil. *Rev Patol Trop* 2016; 45(4): 361-368. http://dx.doi.org/10.5216/rpt. v45i4.44639.

Coklin T, Farber J, Parrington L, Dixon B. Prevalence and molecular characterization of *Giardia duodenalis* and *Cryptosporidium* spp. in dairy cattle in Ontario, Canada. *Vet Parasitol* 2007; 150(4): 297-305. http://dx.doi.org/10.1016/j.vetpar.2007.09.014. PMid:17964724.

Cosendey RIJ, Fiuza VRS, Teixeira CS, Oliveira FCR. Freqüência de oocistos de coccídios do gênero *Cryptosporidium* em ovinos no estado do Rio de Janeiro. *Rev Bras Saúde Prod Anim* 2008; 9(4): 687-669.

Couto MCM, Lima MF, Bomfim TCB. New *Cryptosporidium parvum* subtypes of IIa subfamily in dairy calves from Brazil. *Acta Trop* 2014; 130: 117-122. http://dx.doi.org/10.1016/j.actatropica.2013.11.002. PMid:24239750.

Ederli BB, Carvalho CB, Sales LG. Ocorrência da infecção por *Cryptosporidium* em bezerros na microrregião de Campos dos Goytacazes no norte do Estado do Rio de Janeiro, Brasil. *Rev Bras Parasitol Vet* 2004; 13(2): 45-48.

Fayer R, Santín M, Trout JM. *Cryptosporidium ryanae* n. sp. (Apicomplexa: Cryptosporidiidae) in cattle (*Bos taurus*). *Vet Parasitol* 2008; 156(3-4): 191-198. http://dx.doi.org/10.1016/j.vetpar.2008.05.024. PMid:18583057.

Feitosa FLF, Shimamura GM, Roberto T, Meireles MV, Nunes CM, Ciarlini PC, et al. Prevalência de Criptosporidiose em bezerros na região de Araçatuba, Estado de São Paulo, Brasil. *Cienc Rural* 2004; 34(1): 189-193. http://dx.doi.org/10.1590/S0103-84782004000100029.

Féres FC, Lombardi AL, Carvalho MPP, Mendes LCN, Peiró JR, Cadioli FA, et al. Ocorrência e caracterização molecular de *Cryptosporidium* em cordeiros. *Arq Bras Med Vet Zootec* 2009; 61(4): 1002-1005. http://dx.doi. org/10.1590/S0102-09352009000400033.

Fiuza VR, Cosendey RI, Frazão-Teixeira E, Santín M, Fayer R, Oliveira FC. Molecular characterization of *Cryptosporidium* in Brazilian sheep. *Vet Parasitol* 2011; 175(3-4): 360-362. http://dx.doi.org/10.1016/j. vetpar.2010.10.036. PMid:21075526.

Fujii KY, Dittrich JR, Castro EA, Almeida JC. Ocorrência de *Cryptosporidium* spp em dois centros de treinamento de equinos de Curitiba, Paraná. *Braz J Vet Res Anim Sci* 2014; 51(2): 118-121. http://dx.doi.org/10.11606/ issn.2318-3659.v51i2p118-121.

Garcia AM, Lima JD. Frequência do *Cryptosporidium* em bezerros lactentes de rebanhos leiteiros de Minas Gerais. *Arq Bras Med Vet Zootec* 1993; 45(2): 193-198.

Garcia AM, Lima JD. Prevalência de *Cryptosporidium* spp em rebanhos leiteiros de Pará de Minas (M.G.) e sua relação com práticas de manejo. *Rev Bras Parasitol Vet* 1994; 3(1): 23-28.

Gow S, Waldner C. An examination of the prevalence of and risk factors for shedding of *Cryptosporidium* spp. and *Giardia* spp. in cows and calves

from western Canadian cow – calf herds. *Vet Parasitol* 2006; 137(1-2): 50-61. http://dx.doi.org/10.1016/j.vetpar.2005.05.071. PMid:16472921.

Heckler RP, Borges DGL, Bacha FB, Onizuka MKV, Teruya LS, Neves JPL, et al. First genetic identification of *Cryptosporidium parvum* subtype IIaA14G2R1in beef cattle in Brazil. *Prev Vet Med* 2015; 121(3-4): 391-394. http://dx.doi.org/10.1016/j.prevetmed.2015.08.016. PMid:26342791.

Holsback L, Luppi PAR, Silva CS, Negrão GK, Conde G, Gabriel HV, et al. Anthelmintic efficiency of doramectin, fenbendazole, and nitroxynil, in combination or individually, in sheep worm control. *Rev Bras Parasitol Vet* 2016; 25(3): 353-358. http://dx.doi.org/10.1590/ S1984-29612016025. PMid:27096532.

Holsback L, Marquez ES, Meneghel PP. Resistência parasitária de helmintos gastrointestinais e avaliação dos parâmetros hematológicos de ovinos no norte do Paraná. *Rev Bras Med Vet* 2013; 35(1): 76-84.

Holsback L, Silva MA, Patelli THC, Jesus AP, Sanches JR. Resistance of *Haemonchus, Cooperia, Trichostrongylus*, and *Oesophagostomum* to ivermectin in dairy cattle in Paraná. *Semina: Ciênc Agrár* 2015; 36(3): 2031-2036. http://dx.doi.org/10.5433/1679-0359.2015v36n3Supl1p2031.

Kvác M, Vitovec J. Prevalence and pathogenicity of *Cryptosporidium* andersoni in one herd of beef cattle. *J Vet Med B Infect Dis Vet Public Health* 2003; 50(9): 451-457. http://dx.doi.org/10.1046/j.0931-1793.2003.00701.x. PMid:14633200.

Levine ND. Phylum II Apicomplexa. In: Lee JJ, Hutner SH, Bovee EC, editors. *An illustrated guide to the protozoa*. Lawrence: Allen Press; 1985. p. 322-374.

Lima RCA, Aquino MCC, Inácio SV, Viol MA, Zucatto AS, Silveira L No, et al. Caracterização molecular de *Cryptosporidium* spp. em bezerros (*Bos taurus e Bos indicus*) no município de Formiga, Minas Gerais -Brasil. *Semina: Ciênc Agrár* 2013; 34(6): 3747-3754. http://dx.doi. org/10.5433/1679-0359.2013v34n6Supl2p3747.

Majewska AC, Werner A, Sulima P, Luty T. Prevalence of *Cryptosporidium* in sheep and goats bred on five farms in west-central region of Poland. *Vet Parasitol* 2000; 89(4): 269-275. http://dx.doi.org/10.1016/S0304-4017(00)00212-0. PMid:10799840.

Martinez I, Belda FM No. Contribution to the laboratory diagnosis of human cryptosporidiosis. *Rev Inst Med Trop São Paulo* 2001; 43(2): 79-82. http://dx.doi.org/10.1590/S0036-46652001000200005. PMid:11340480.

Morgan UM, Pallant L, Dwyer BW, Forbes DA, Rich G, Thompson RCA. Comparison of PCR and microscopy for detection of *Cryptosporidium parvum* in human fecal specimens: clinical trial. *J Clin Microbiol* 1998; 36(4): 995-998. PMid:9542924.

Nakamura AA, Simões DC, Antunes RG, Silva DC, Meireles MV. Molecular characterization of *Cryptosporidium* spp. from fecal samples of birds kept in captivity in Brazil. *Vet Parasitol* 2009; 166(1-2): 47-51. http://dx.doi.org/10.1016/j.vetpar.2009.07.033. PMid:19683397.

Navarro IT, Kano FS, Ogawa L, Freire RL, Vidotto O. Ocorrência de *Cryptosporidium* spp em cães com diarréia atendidos no Hospital Veterinário da Universidade Estadual de Londrina, PR, Brasil. *Semina: Ciênc Agrár* 1997; 18(1): 23-25.

Paz e Silva FM, Lopes RS, Araújo-Júnior JP. Identification of *Cryptosporidium* species and genotypes in dairy cattle in Brazil. *Rev Bras Parasitol Vet* 2013; 22(1): 22-28.

Paz e Silva FM, Lopes RS, Bresciani KD, Amarante AF, Araujo JP Jr. High occurrence of *Cryptosporidium ubiquitum* and *Giardia duodenalis* genotype E in sheep from Brazil. *Acta Parasitol* 2014; 59(1): 193-196. http://dx.doi.org/10.2478/s11686-014-0223-5. PMid:24570068.

Ralston BJ, Cockwill CL, Guselle NJ, Van Herk FH, McAllister TA, Olson ME. Prevalence of *Giardia* and *Cryptosporidium andersoni* and their effects on performance in feedlot beef cattle. *Can J Anim Sci* 2003; 83(1): 153-159. http://dx.doi.org/10.4141/A01-001.

Rekha HKM, Puttalakshmamma GC, D'Souza PE. Comparison of different diagnostic techniques for the detection of cryptosporidiosis in bovines. *Vet World* 2016; 9(2): 211-215. http://dx.doi.org/10.14202/ vetworld.2016.211-215. PMid:27051211.

Robinson G, Thomas AL, Daniel RG, Hadfield SJ, Elwin K, Chalmers RM. Sample prevalence and molecular characterisation of *Cryptosporidium andersoni* within a dairy herd in the United Kingdom. *Vet Parasitol* 2006; 142(1-2): 163-167. http://dx.doi.org/10.1016/j.vetpar.2006.06.031. PMid:16908101.

Santín M, Trout JM, Fayer R. Prevalence and molecular characterization of *Cryptosporidium* and *Giardia* species and genotypes in sheep in Maryland. *Vet Parasitol* 2007; 146(1-2): 17-24. http://dx.doi.org/10.1016/j. vetpar.2007.01.010. PMid:17335979.

Santín M, Trout JM, Fayer R. A longitudinal study of cryptosporidiosis in dairy cattle from birth to 2 years of age. *Vet Parasitol* 2008; 155(1-2): 15-23. http://dx.doi.org/10.1016/j.vetpar.2008.04.018. PMid:18565677.

Sevá AP, Funada MR, Souza SO, Nava A, Richtzenhain LJ, Soares RM. Occurrence and molecular characterization of *Cryptosporidium* spp. isolated from domestic animals in a rural area surrounding Atlantic dry forest fragments in Teodoro Sampaio municipality, State of São Paulo, Brazil. *Rev Bras Parasitol Vet* 2010; 19(4): 249-253. http://dx.doi.org/10.1590/ S1984-29612010000400011. PMid:21184703.

Silva FM, Lopes RS, Araújo-Junior JP. Identification of *Cryptosporidium* species and genotypes in dairy cattle in Brazil. *Rev Bras Parasitol Vet* 2013; 22(1): 22-28. http://dx.doi.org/10.1590/S1984-29612013005000010. PMid:23538500.

Silva-Junior FA, Carvalho AHO, Rocha CMBM, Guimaráes AM. Fatores de risco associados à infecção por *Cryptosporidium* spp e *Giardia duodenalis* em bovinos leiteiros na fase de cria e recria na mesorregião do Campo das Vertentes de Minas Gerais. *Pesq Vet Bras* 2011; 31(8): 690-696. http://dx.doi.org/10.1590/S0100-736X2011000800010.

Snak A, Garcia FG, Delgado LES, Osaki SC. Occurrence of *Cryptosporidium* spp. in wild animals living in the Cascavel city park, Paraná, Brazil. *Semina:*

Ciênc Agrár 2015; 36(6): 4323-4332. http://dx.doi.org/10.5433/1679-0359.2015v36n6Supl2p4323.

Snak A, Smiderle FR, Fernandes NLM, Lara AA, Garcia FG, Ogawa L, et al. Ocurrence and molecular characterization of *Cryptosporidium* sp. in sheep. *Semina: Ciênc Agrár* 2017; 38(4): 1917-1924. http://dx.doi. org/10.5433/1679-0359.2017v38n4p1917.

Tahvildar-Biderouni F, Salehi N. Detection of *Cryptosporidium* infection by modified ziehl-neelsen and PCR methods in children with diarrheal samples in pediatric hospitals in Tehran. *Gastroenterol Hepatol Bed Bench* 2014; 7(2): 125-130. PMid:24834304.

Toledo RS, Martins FDC, Ferreira FP, Almeida JC, Ogawa L, Santos HLEPL, et al. *Cryptosporidium* spp. and *Giardia* spp. in feces and water and the associated exposure factors on dairy farms. *PLoS One* 2017; 12(4): e0175311. http://dx.doi.org/10.1371/journal.pone.0175311. PMid:28403147.

Villacorta I, Ares-Mazas E, Lorenzo MJ. *Cryptosporidium parvum* in cattle, sheep and pigs in Galicia (N.W. Spain). *Vet Parasitol* 1991; 38(2-3): 249-252. http://dx.doi.org/10.1016/0304-4017(91)90134-H. PMid:1858293.

Wells B, Shaw H, Hotchkiss E, Gilray J, Ayton R, Green J, et al. Prevalence, species identification and genotyping *Cryptosporidium* from livestock and deer in a catchment in the Cairngorms with a history of a contaminated public water supply. *Parasit Vectors* 2015; 8(1): 66. http:// dx.doi.org/10.1186/s13071-015-0684-x. PMid:25650114.

Xiao L, Escalante L, Yang C, Sulaiman I, Escalante AA, Montali RJ, et al. Phylogenetic analysis of *Cryptosporidium* parasites based on the smallsubunit rRNA gene locus. *Appl Environ Microbiol* 1999; 65(4): 1578-1583. PMid:10103253.

Xiao L, Herd RP, Rings DM. Diagnosis of *Cryptosporidium* on a sheep farm with neonatal diarrhea by immunofluorescence assays. *Vet Parasitol* 1993; 47(1-2): 17-23. http://dx.doi.org/10.1016/0304-4017(93)90172-J. PMid:8493764.

Zucatto AS, Aquino MCC, Inácio SV, Figueiredo RN, Pierucci JC, Perri SHV, et al. Molecular characterisation of *Cryptosporidium* spp. in lambs in the South Central region of the State of São Paulo. *Arq Bras Med Vet Zootec* 2015; 67(2): 441-446. http://dx.doi.org/10.1590/1678-7067.