

'*Candidatus Mycoplasma haemoalbiventris*' and tick-borne pathogens screening in white-eared opossums (*Didelphis albiventris*) from Curitiba and Foz do Iguaçu Cities, Paraná State, southern Brazil

'*Candidatus Mycoplasma haemoalbiventris*' e triagem para patógenos transmitidos por carrapatos em gambás de orelha branca (*Didelphis albiventris*) das cidades de Curitiba e Foz do Iguaçu, Estado do Paraná, sul do Brasil

Renata Prestes Antonangelo de Oliveira¹; Flávia Carolina Meira Collere¹; Larissa Dantas Roeder Ferrari¹; Vanessa dos Santos Coradi¹; Nathália de Albuquerque Soares¹; André de Souza Leandro²; Wagner Fabiano de Oliveira²; Sandro Roberto Galvão²; Rosinei Kafka²; Robson Michael Delai³; Rafaella Martini⁴; André Saldanha¹; Leonardo Pereira dos Santos¹; Zalmir Silvino Cubas⁵; Rogério Ribas Lange⁴; Thállitha Samih Wischral Jayme Vieira^{1,6*} 

¹Laboratório de Doenças Transmitidas por Vetores, Departamento de Medicina Veterinária, Universidade Federal do Paraná – UFPR, Curitiba, PR, Brasil

²Unidade de Vigilância em Zoonoses, Secretaria Municipal de Saúde, Foz do Iguaçu, PR, Brasil

³Laboratório de Saúde Única, Centro de Medicina Tropical das Três Fronteiras, Fundação Itaiguapy, Instituto de Ensino e Pesquisa, Foz do Iguaçu, PR, Brasil

⁴Departamento de Medicina Veterinária, Universidade Federal do Paraná – UFPR, Curitiba, PR, Brasil

⁵Médico Veterinário Autônomo, Foz do Iguaçu, PR, Brasil

⁶Global One Health initiative – GOHi, The Ohio State University, Columbus, OH, USA

How to cite: Oliveira RPA, Collere FCM, Ferrari LDR, Coradi VS, Soares NA, Leandro AS, et al. '*Candidatus Mycoplasma haemoalbiventris*' and tick-borne pathogens screening in white-eared opossums (*Didelphis albiventris*) from Curitiba and Foz do Iguaçu Cities, Paraná State, southern Brazil. *Braz J Vet Parasitol* 2021; 30(4): e009721. <https://doi.org/10.1590/S1984-29612021072>

Abstract

Hemoplasmas are epierthrocytic bacteria that infect mammals. '*Candidatus Mycoplasma haemoalbiventris*' was detected in white-eared opossums (*Didelphis albiventris*) from southern and central-western Brazil. The present study aimed at: i) screening opossums for tick-borne (TBP) pathogens (Piroplasmida and Anaplasmataceae) and ii) detecting and characterizing hemoplasma species infecting opossums from Curitiba and Foz do Iguaçu cities in the Paraná State, southern Brazil. Thirty blood samples from white-eared opossums were evaluated by PCR assays. Animals were not infested by ectoparasites. The mammalian endogenous *gapdh* gene was consistently amplified in all samples. All opossums tested negative for *Theileria/Babesia* spp. and *Ehrlichia/Anaplasma* spp. by PCR based on 18S rRNA and 16S rRNA genes, respectively. A genus-specific PCR assay based on the 16S rRNA gene of hemoplasmas showed that three/13 (23.08%; CI 95%: 8.18-50.26%) opossums from Foz do Iguaçu were positive for hemotropic *Mycoplasma* sp. All opossums from Curitiba tested negative for hemoplasmas. Sequencing of both the 16S and 23S rRNA genes revealed that the animals were infected by '*Ca. M. haemoalbiventris*'. Although '*Ca. M. haemoalbiventris*' is prevalent in opossums in Brazil, clinical signs associated with its infection and its putative vectors remain unknown.

Keywords: Marsupials, hemotropic mycoplasmas, hemoplasmas.

Resumo

Hemoplasmas são bactérias epieritrocíticas que infectam mamíferos. '*Candidatus Mycoplasma haemoalbiventris*' foi detectado previamente em gambás-de-orelha-branca (*Didelphis albiventris*) das regiões sul e centro-oeste do

Received May 26, 2021. Accepted July 13, 2021.

*Corresponding author: Rafael Felipe da Costa Vieira. E-mail: rvieira@ufpr.br



This is an Open Access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Brasil. O presente estudo objetivou: i) triar os gambás para as doenças transmitidas por carrapatos (Piroplasmida e Anaplasmataceae); e ii) detectar e caracterizar as espécies de hemoplasma que infectam gambás nas cidades de Curitiba e Foz do Iguaçu, no Estado do Paraná, sul do Brasil. Trinta amostras de sangue de gambás-de-orelha-branca foram analisadas por PCR. Os animais não estavam infestados por ectoparasitos. O gene endógeno de mamífero *gapdh* foi amplificado em todas as amostras. Todos os gambás testaram negativos para *Theileria/Babesia* spp. e *Ehrlichia/Anaplasma* spp. por PCR, respectivamente, para os genes 18S rRNA e 16S rRNA. Uma PCR gene-específica, baseada no gene 16S rRNA de hemoplasmas, mostrou que três/13 (23,08%; CI 95%: 8,18-50,26%) gambás de Foz do Iguaçu foram positivos para *Mycoplasma* sp. hemotrópico. Todos os gambás de Curitiba testaram negativos para hemoplasmas. O sequenciamento de fragmentos dos genes 16S e 23S rRNA revelou que os animais estavam infectados pelo 'Ca. M. haemoalbiventris'. Embora 'Ca. M. haemoalbiventris' seja prevalente em gambás no Brasil, os sinais clínicos associados à infecção e os prováveis vetores permanecem desconhecidos.

Palavras-chave: Marsupiais, micoplasma hemotrópico, hemoplasmas.

Opossums are synanthropic marsupials belonging to the genus *Didelphis*. In South America, four species have been reported: *D. albiventris*, *D. aurita*, *D. imperfecta*, and *D. marsupialis* (Nascimento & Horta, 2014). Due to their circulation in urban and rural environments, opossums are potential hosts, suggested reservoirs and/or potential amplifiers of infectious agents (e.g., *Trypanosoma cruzi*, *Leishmania infantum*, *Rickettsia rickettsii*) (Horta et al., 2009; Carreira et al., 2012; Zecca et al., 2020).

In Brazil, tick-borne pathogens (TBP) have been reported in *Didelphis* spp. from different regions (Melo et al., 2016; Soares et al., 2017; Guimarães et al., 2019). This includes a novel ehrlichial agent, *Ehrlichia* sp. strain Natal, detected in white-eared opossum (*D. albiventris*) from the northeastern region of the country (Lopes et al., 2018).

Hemotropic mycoplasmas (hemoplasmas) are Gram-negative bacteria that attach to the surface of red blood cells from mammals and may cause hemolytic anemia. Two hemotropic *Mycoplasma* species have been described in *Didelphis*, namely 'Candidate Mycoplasma haemodidelphis' (Messick et al., 2002) and 'Candidate Mycoplasma haemoalbiventris' (Pontarolo et al., 2021). Due to the proximity of human dwellings and human-animal interactions, monitoring ticks and the health status of opossums is a public health concern. Therefore, this study aimed at investigating the occurrence of TBP and hemotropic mycoplasmas in free-ranging opossums from the Curitiba and Foz do Iguaçu cities of the Paraná State, southern Brazil.

This study was approved by the Ethics Committee for Animal Experimentation and Animal Welfare at the Universidade Federal do Paraná, Brazil (protocol number 053/2018). Animal and laboratory procedures were approved and performed under the regulations of the Chico Mendes Institute for Biodiversity Conservation (ICMBio, protocol number 63433-3).

The study was carried out in the Curitiba (25° 26' 27" S 49° 16' 36" W) and Foz do Iguaçu cities (25° 32' 45" S 54° 35' 07" W). Curitiba, the capital of Paraná State, is located in the south-central region and has a humid subtropical highland climate (Köppen: Cfb) with an average temperature of 17.4 °C. Foz do Iguaçu lies in the extreme west of the Paraná State, on the border of Brazil, Argentina, and Paraguay, in the Atlantic rainforest biome. Foz do Iguaçu is internationally recognized as a protected area with diverse fauna (Valente et al., 2019) and has a humid subtropical climate (Köppen: Cfa) with rainfall throughout the year and an average temperature of 22.1 °C.

A total of 30 white-eared opossums (*D. albiventris*) were evaluated. In Foz do Iguaçu, 13 white-eared opossums were captured using Tomahawk wire mesh traps baited with fruit. Sampling was performed between April and December 2019 per the spontaneous demand of the Zoonoses Surveillance Center, based on the report of the occurrence of opossums in human dwellings. In Curitiba City, 17 white-eared opossums were referred to the Veterinary Teaching Hospital, Universidade Federal do Paraná, Parana State, southern Brazil, between June 2018 and December 2020. Samples from Curitiba were also collected per the spontaneous demand based on the report of the occurrence of opossums in human dwellings.

After using chemical restraints (xylazine (4.0 mg/kg) and ketamine (20 mg/kg), the animals were individually identified and visually inspected for ectoparasites (ticks and fleas). Subsequently, EDTA-blood samples were collected by caudal venipuncture and stored at -20 °C until molecular analysis. Post sample collection, the animals were monitored until complete recovery from the chemical restraints and later released (Massini et al., 2019).

DNA was extracted from 200 µL of blood using a commercially available kit (Illustra™ Blood Genomic Prep Mini Spin Kit, GE Healthcare Life Sciences, Little Chalfont, UK), according to the manufacturer's instructions. Ultra-pure water was used as a negative control in parallel to monitor cross-contamination.

To monitor DNA extraction, PCR for the mammalian endogenous gene glyceraldehyde-3-phosphate dehydrogenase (*gapdh*) was performed in all samples (Birkenheuer et al., 2003). Conventional PCR was used to screen samples for hemotropic *Mycoplasma* spp. genus-specific primers targeting a portion (900 bp) of the 16S rRNA gene of hemotropic *Mycoplasma* spp. (Hoelzle et al., 2011; Machado et al., 2017). Opossum DNA samples that tested positive in the PCR assay based on the 16S rRNA gene were subjected to a genus-specific PCR assay targeting a fragment (800 bp) of the 23S rRNA gene of hemoplasmas (Mongruel et al., 2020). Nuclease-free water and *Mycoplasma ovis* DNA from a naturally infected goat blood sample were used as the negative and positive controls, respectively, in both PCR assays. Additionally, DNA samples were also tested by PCR assays targeting a fragment (551 bp) of the 18S rRNA gene of *Theileria/Babesia* spp. (Almeida et al., 2012) and a fragment (349 bp) of the 16S rRNA gene of *Ehrlichia/Anaplasma* spp. (Parola et al., 2000). *Babesia vogeli* and *Ehrlichia canis* DNA obtained from naturally infected dogs were used as positive controls, and nuclease-free water was used as a negative control (Vieira et al., 2013). The amplified PCR products were subjected to gel electrophoresis in 1.5% agarose gels for 1 hour at 100 V, followed by SYBR safe staining (6 µg/mL; SYBR® Safe DNA Gel Stain, Invitrogen, CA, USA), and viewed under a UV light transilluminator at a wavelength of 312 nm.

The amplicons of the 16S rRNA (900 bp) and 23S rRNA (800 bp) obtained from three and two *Mycoplasma* spp.-positive samples were from Foz do Iguaçu. They were sequenced in both directions using the Sanger method. Three nucleotide sequences of the 16S rRNA gene and two sequences of 23S rRNA gene of hemotropic *Mycoplasma* sp. amplified herein were submitted to the GenBank® database (accession numbers: MW703800, MW703801, MW703802, and MW694786 and MW694787, respectively).

The consensus sequences were subjected to multiple alignment with the sequences selected from GenBank® using MAFFT available on the GUIDANCE2 server for each gene. The best-fit model of nucleotide substitution was determined using jModeltest v.2.1.10 and was set as GTR+I+G for 16S rRNA and TrN+G for 23S rRNA genes, based on the Akaike Information Criterion (AIC), respectively. Each Bayesian reconstruction was performed in Beast 1.10.4 with three independent runs of 100 million MCMC steps sampled at every 10,000 trees, 10% of the burn-in. The phylogenetic tree was visualized with FigTree software version 1.4.4 and the final layout was done with Inkscape version 0.92.2.

Opossums from both cities were not infested by ectoparasites (ticks and fleas) at the time of sampling. The mammalian endogenous *gapdh* gene was consistently amplified in all samples. All opossums tested negative for *Theileria/Babesia* spp. and *Ehrlichia/Anaplasma* spp. by PCR. While all opossums from Curitiba tested negative for hemoplasmas, three out of 13 (23.08%; CI 95%: 8.18-50.26%) animals from Foz do Iguaçu were positive for hemotropic *Mycoplasma* spp., as observed via PCR.

Sequencing of the 16S rRNA fragment showed 100% identity to '*Ca. M. haemoalbiventris*' (MH158514, MH158515, MN423256, MN423258-MN423260, and MT170012-MT170016) detected in white-eared opossums from Brazil; on the other hand, sequencing of the 23S rRNA fragment showed 99.74-100% identity '*Ca. M. haemoalbiventris*' (MN442081-MN442085) detected in white-eared opossums from Brazil. Phylogenetic analysis of the 16S and 23S rRNA gene fragments confirmed the white-eared opossum were infected by '*Ca. M. haemoalbiventris*' (Figures 1 and 2).

In the present study, 23.08% opossums from Foz do Iguaçu (Paraná, southern Brazil) were positive for '*Ca. M. haemoalbiventris*', while all the animals from Curitiba City tested negative. Previous studies have found higher hemoplasma prevalence rates (ranging from 32.5-87.5%) in white-eared opossums from different geographic regions and biomes (Atlantic Forest and Cerrado) of Brazil (Massini et al., 2019; Gonçalves et al., 2020; Pontarolo et al., 2021). Although Curitiba City is also located in the Atlantic rainforest biome, we hypothesize that its altitude (940 m a.s.l.) and annual average temperature (17.4 °C) may influence habits and behavior of the opossums, and thus, their exposure to hemoplasmas, which may explain the negative results found herein. Additionally, differences in the prevalence of hemoplasma between studies may have not be due to the diagnostic test used, since to date, all studies on the detection of hemoplasmas in Brazilian opossums have used conventional PCR assays as diagnostic method (Massini et al., 2019; Gonçalves et al., 2020; Pontarolo et al., 2021).

Herein, the animals were not infested with ticks and fleas at the time of sampling. However, hemoplasma-positive white-eared opossums from the Maringá municipality (state of Paraná) (Massini et al., 2019) and Campo Grande municipality (state of Mato Grosso do Sul) (Gonçalves et al., 2020) were infested by *Amblyomma dubitatum* ticks. On the other hand, opossums from the Canoinhas municipality (state of Santa Catarina) were infested with

Ctenocephalides felis fleas (Pontarolo et al., 2021). Previous studies have failed to detect hemotropic *Mycoplasma* spp. in *A. dubitatum* ticks infesting 'Ca. M. haemoalbiventris'-positive opossums from Brazil (Gonçalves et al., 2020), and canine hemoplasmas in *Rhipicephalus sanguineus* sensu lato ticks in dogs from an animal shelter in Turkey (Aktas & Ozubek, 2017). High hemoplasma prevalence rates have been found in tropical regions, favoring the establishment of arthropod vectors for these bacteria. Additionally, hemotropic *Mycoplasma* spp. have been detected in *Amblyomma sculptum* ticks infesting hemoplasma-positive capybaras (*Hydrochoerus hydrochaeris*) from central-western Brazil (Gonçalves et al., 2020). However, the lack of experimental studies on hemoplasma transmission, due to its uncultivable status *in vitro*, precludes robust evidence to implicate the involvement of ticks in the epidemiological cycle of this group of bacteria. Finally, *A. dubitatum* ticks are widespread and have already been found to infest opossums from Curitiba and Foz do Iguaçu cities (Valente et al., 2020). Based on the above evidence, we hypothesize that ticks from this group may have previously bitten animals evaluated in the present study, although we cannot associate the low hemoplasma infection rate found to the absence of ticks.

TBP are of great concern worldwide. In Brazil, *Ehrlichia* spp., *Anaplasma* spp., and *Babesia* spp. have been reported to infect domestic and wild animals (Soares et al., 2017; André, 2018). A novel ehrlichial agent, *Ehrlichia* sp. strain Natal, has been detected in the northeastern region of the country (Lopes et al., 2018). A putative novel genotype of *Ehrlichia* sp. has been detected in big-eared opossums (*D. aurita*) from southeastern Brazil (Guimarães et al., 2019). Morphological identification of piroplasmids (previously named as *Nuttallia brasiliensis*/*Theileria brasiliensis*/*Babesia ernestoi*, and currently *Babesia brasiliensis*) in marsupials circulating in Brazil was performed virtually 100 years ago (Regendanz & Kikuth, 1928). Additionally, a novel piroplasmid closely related to a *Babesia* sp. detected in *Monodelphis domestica* opossums from the Brazilian Pantanal has been found in *D. marsupialis* opossums from the Brazilian Amazon (Soares et al., 2017), and another Piroplasmida genotype was reported in two/31 (6.45%) *D. marsupialis* trapped in Sinop municipality, State of Mato Grosso (Colle et al., 2019). In the present study, all white-eared opossums tested negative for *Theileria/Babesia* spp. and *Ehrlichia/Anaplasma* spp. Although neither clinical signs of infection nor tick vectors have been established for the novel pathogens described in opossums from Brazil, further studies should focus on evaluating this group of marsupials to better understand and characterize the pathogens found.

In conclusion, 'Candidate *Mycoplasma haemoalbiventris*' has been detected in white-eared opossums from the Foz do Iguaçu city (Paraná state, southern Brazil), while the animals from Curitiba city tested negative. However, the clinical signs associated with the infection caused by this organism and its putative vectors remain unknown. All animals from both cities tested negative for *Theileria/Babesia* spp. and *Ehrlichia/Anaplasma* spp.

Acknowledgments

This research was funded by Universidade Federal do Paraná (Grant no. 23075.058259/2020-23). This study is part of a PhD degree for Renata Antonangelo at the Universidade Federal do Paraná. Flávia Collere, Rafaella Martini and André Saldanha were sponsored by a fellowship from the the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) at the time of research. Vanessa Coradi and Nathália Soares were sponsored by a fellowship from the Brazilian National Council of Scientific and Technological Development (CNPq). CNPq also provided a fellowship of research productivity (PQ) to Dr. Rafael F.C. Vieira (CNPq - 313161/2020-8).

References

- Aktas M, Ozubek S. Molecular survey of haemoplasmas in shelter dogs and associations with *Rhipicephalus sanguineus sensu lato*. *Med Vet Entomol* 2017; 31(4): 457-461. <http://dx.doi.org/10.1111/mve.12244>. PMID:28685834.
- Almeida AP, Marcili A, Leite RC, Nieri-Bastos FA, Domingues LN, Martins JR, et al. *Coxiella* symbiont in the tick *Ornithodoros rostratus* (Acari: argasidae). *Ticks Tick Borne Dis* 2012; 3(4): 203-206. <http://dx.doi.org/10.1016/j.ttbdis.2012.02.003>. PMID:22480930.
- André MR. Diversity of *Anaplasma* and *Ehrlichia/Neoehrlichia* agents in terrestrial wild carnivores worldwide: implications for human and domestic animal health and wildlife conservation. *Front Vet Sci* 2018; 5: 293. <http://dx.doi.org/10.3389/fvets.2018.00293>. PMID:30533417.
- Birkenheuer AJ, Levy MG, Breitschwerdt EB. Development and evaluation of a seminested PCR for detection and differentiation of *Babesia gibsoni* (Asian genotype) and *B. canis* DNA in canine blood samples. *J Clin Microbiol* 2003; 41(9): 4172-4177. <http://dx.doi.org/10.1128/JCM.41.9.4172-4177.2003>. PMID:12958243.
- Carreira JCA, Silva AVM, Pita Pereira D, Brazil RP. Natural infection of *Didelphis aurita* (Mammalia: Marsupialia) with *Leishmania infantum* in Brazil. *Parasit Vectors* 2012; 5(1): 111. <http://dx.doi.org/10.1186/1756-3305-5-111>. PMID:22676324.

- Colle AC, Mendonça RFB, Maia MO, Freitas LC, Witter R, Marcili A, et al. Molecular survey of tick-borne pathogens in small mammals from Brazilian Amazonia. *Rev Bras Parasitol Vet* 2019; 28(4): 592-604. <http://dx.doi.org/10.1590/s1984-29612019086>. PMID:31800885.
- Gonçalves LR, Herrera HM, Nantes WAG, Santos FM, Porfírio GEO, Barreto WGT, et al. Genetic diversity and lack of molecular evidence for hemoplasma cross-species transmission between wild and synanthropic mammals from Central-Western Brazil. *Acta Trop* 2020; 203: 105303. <http://dx.doi.org/10.1016/j.actatropica.2019.105303>. PMID:31857081.
- Guimarães A, Raimundo JM, Silva AT, Carpintero FM, Pires JR, Benevenuto JL, et al. Detection of a putative novel genotype of *Ehrlichia* sp. from opossums (*Didelphis aurita*) from Brazil. *Rev Bras Parasitol Vet* 2019; 28(1): 140-144. <http://dx.doi.org/10.1590/s1984-296120180068>. PMID:30427523.
- Hoelzle K, Winkler M, Kramer MM, Wittenbrink MM, Dieckmann SM, Hoelzle LE. Detection of *Candidatus Mycoplasma haemobos* in cattle with anaemia. *Vet J* 2011; 187(3): 408-410. <http://dx.doi.org/10.1016/j.tvjl.2010.01.016>. PMID:20188610.
- Horta MC, Moraes-Filho J, Casagrande RA, Saito TB, Rosa SC, Ogrzewalska M, et al. Experimental infection of opossums *Didelphis aurita* by *Rickettsia rickettsii* and evaluation of the transmission of the infection to ticks *Amblyomma cajennense*. *Vector Borne Zoonotic Dis* 2009; 9(1): 109-118. <http://dx.doi.org/10.1089/vbz.2008.0114>. PMID:18945194.
- Lopes MG, Muñoz-Leal S, de Lima JTR, Fournier GFSR, Acosta ICL, Martins TF, et al. Ticks, rickettsial and ehrlichial infection in small mammals from Atlantic forest remnants in northeastern Brazil. *Int J Parasitol Parasites Wildl* 2018; 7(3): 380-385. <http://dx.doi.org/10.1016/j.ijppaw.2018.10.001>. PMID:30370217.
- Machado CAL, Vidotto O, Conrado FO, Santos NJR, Valente JDM, Barbosa IC, et al. *Mycoplasma ovis* infection in goat farms from northeastern Brazil. *Comp Immunol Microbiol Infect Dis* 2017; 55: 1-5. <http://dx.doi.org/10.1016/j.cimid.2017.08.004>. PMID:29127988.
- Massini PF, Drozino RN, Otomura FH, Mongrue ACB, Valente JDM, Toledo MJO, et al. Detection of Hemotropic *Mycoplasma* sp. in white-eared opossums (*Didelphis albiventris*) from Southern Brazil. *Rev Bras Parasitol Vet* 2019; 28(4): 797-801. <http://dx.doi.org/10.1590/s1984-29612019058>. PMID:31390439.
- Melo ALT, Aguiar DM, Spolidorio MG, Yoshinari NH, Matushima ER, Labruna MB, et al. Serological evidence of exposure to tick-borne agents in opossums (*Didelphis* spp.) in the state of São Paulo, Brazil. *Rev Bras Parasitol Vet* 2016; 25(3): 348-352. <http://dx.doi.org/10.1590/S1984-29612016028>. PMID:27276663.
- Messick JB, Walker PG, Raphael W, Berent L, Shi X. '*Candidatus Mycoplasma haemodidelphidis*' sp. nov., '*Candidatus Mycoplasma haemolamae*' sp. nov. and *Mycoplasma haemocanis* comb. nov., haemotrophic parasites from a naturally infected opossum (*Didelphis virginiana*), alpaca (*Lama pacos*) and dog (*Canis familiaris*): phylogenetic and secondary structural relatedness of their 16S rRNA genes to other mycoplasmas. *Int J Syst Evol Microbiol* 2002; 52(3): 693-698. <http://dx.doi.org/10.1099/00207713-52-3-693>. PMID:12054227.
- Mongrue ACB, Spanhol VC, Valente JDM, Porto PP, Ogawa L, Otomura FH, et al. Survey of vector-borne and nematode parasites involved in the etiology of anemic syndrome in sheep from Southern Brazil. *Rev Bras Parasitol Vet* 2020; 29(3): e007320. <http://dx.doi.org/10.1590/s1984-29612020062>. PMID:32935770.
- Nascimento CC, Horta MC. Didelphimorphia (Gambá, Cuíca). In: Cubas ZS, Silva JCR., Catão-Dias JL, editors. *Tratado de animais selvagens: medicina veterinária*. 2. ed. São Paulo: Roca; 2014. (vol. 1).
- Parola P, Roux V, Camicas JL, Baradj I, Brouqui P, Raoult D. Detection of ehrlichiae in African ticks by polymerase chain reaction. *Trans R Soc Trop Med Hyg* 2000; 94(6): 707-708. [http://dx.doi.org/10.1016/S0035-9203\(00\)90243-8](http://dx.doi.org/10.1016/S0035-9203(00)90243-8). PMID:11198664.
- Pontarolo GH, Kühl LF, Pedrassani D, Campos M, Figueiredo FB, Valente JDM, et al. '*Candidatus Mycoplasma haemoalbiventris*', a novel hemoplasma species in white-eared opossums (*Didelphis albiventris*) from Brazil. *Transbound Emerg Dis* 2021; 68(2): 565-572. <http://dx.doi.org/10.1111/tbed.13716>. PMID:32644252.
- Regendanz P, Kikuth W. Sur un parasite du sang des "Quica" (*Metachirus quica*) *Nuttallia brasiliensis* n. sp., et influence de la rate sur les infections latentes du sang. *C R H Seanc Mem Soc Biol* 1928; 98: 1567.
- Soares HS, Marcili A, Barbieri ARM, Minervino AHH, Moreira TR, Gennari SM, et al. Novel piroplasmid and *Hepatozoon* organisms infecting the wildlife of two regions of the Brazilian Amazon. *Int J Parasitol Parasites Wildl* 2017; 6(2): 115-121. <http://dx.doi.org/10.1016/j.ijppaw.2017.05.002>. PMID:28603688.
- Valente JDM, Mongrue ACB, Machado CAL, Chiyo L, Leandro AS, Britto AS, et al. Tick-borne pathogens in carthorses from Foz do Iguaçu City, Paraná State, southern Brazil: A tri-border area of Brazil, Paraguay and Argentina. *Vet Parasitol* 2019; 273: 71-79. <http://dx.doi.org/10.1016/j.vetpar.2019.08.008>. PMID:31446256.
- Valente JDM, Silva PW, Arzua M, Barros-Battesti DM, Martins TF, Silva AM, et al. Records of ticks (Acari: Ixodidae) on humans and distribution of spotted-fever cases and its tick vectors in Paraná State, southern Brazil. *Ticks Tick Borne Dis* 2020; 11(6): 101510. <http://dx.doi.org/10.1016/j.ttbdis.2020.101510>. PMID:32993930.

Vieira TSWJ, Vieira RFC, Nascimento DAG, Tamekuni K, Toledo RS, Chandrashekar R, et al. Serosurvey of tick-borne pathogens in dogs from urban and rural areas from Parana State, Brazil. *Rev Bras Parasitol Vet* 2013; 22(1): 104-109. <http://dx.doi.org/10.1590/S1984-29612013000100019>. PMID:24252955.

Zecca IB, Hodo CL, Slack S, Auckland L, Hamer SA. *Trypanosoma cruzi* infections and associated pathology in urban-dwelling Virginia opossums (*Didelphis virginiana*). *Int J Parasitol Parasites Wildl* 2020; 11: 287-293. <http://dx.doi.org/10.1016/j.ijppaw.2020.03.004>. PMID:32215248.